

J. Biol. Eng. 9: 4; Churgin, M.A, S.K. Jung, C.C. Yu, X. Chen, D.M. Raizen, Fang-Yen C. Elife 2017, May 31: 6; Cini, A., C. Ioriatti, and G. Anfora 2012, Bull. Insectology 65(1): 149-160; Dobzhansky, Th., 1947, Evolution 1: 1-16; Hu, P.J., 2007, Wormbook Aug 8: 1-19; Kiontke, K., and D.H.A. Fitch 2013, Current Biology 23: R862-864; Kiontke, K., and Sudhouse 2006, Wormbook Jan 9: 1-14; Lipson, H.M., and D. Kurman 2013, *Fabricated: The New World of 3D Printing*. Indianapolis, IN: Wiley; Lee, J.C., D.J. Bruck, A.J. Dreves, C. Loriatti, H. Vogt, and P. Baufeld 2011, Pest Manag. Sci. 67: 1349-51; Lee, H., M.K. Choice, D. Lee, H.S. Kim, H. Hwang, H. Kim, S. Park, Y.K. Paik, and J. Lee 2011, Nat. Neurosci. Nov 13: 107-12; makezine.com/comparison/3dprinters/; McDevitt, D., S. McRobert, and J. Fingerut 2017, Dros. Inf. Serv. 100: this issue.



Culture medium for flower-breeding drosophilids.

Larroza, A., G.V. Mello, and J. Cordeiro. Departamento de Ecologia, Zoologia e Genetica; Instituto de Biologia; Universidade Federal de Pelotas, RS, Brazil. E-mail: juliana.cordeiro@ufpel.edu.br

Despite more than 6,800 species of drosophilids are known (Bächli, 2017), very few species are easily maintained in the laboratory and could be used as research models. Among the reasons, it is relatively difficult to keep some lineages, particularly when they have specific developmental requirements. Flower-breeding drosophilids are one of these species that could not be maintained in the laboratory and extremely depends on living flowers. In this context, the recipe presented below was developed to rear flower-breeding drosophilids that do not grow in already described medium, such as the ones from Bizzo *et al.* (2012), Markow and O'Grady (2006), Marques *et al.* (1966), Schmitz (2016), and Vaz *et al.* (2014).

This recipe was developed and has been used in the *Laboratory of Genetic Diversity and Evolution* of *Universidade Federal de Pelotas* (Rio Grande do Sul, Brazil), prepared with flower extracts from *Brugmansia suaveolens* (Solanaceae) and *Ipomoea alba* (Convolvulaceae). However, it can be performed using extracts of other flower species, or even from mushrooms. It proved to be successful for raising *D. bromelioides*-like species (new species not yet properly described, named 'tipo III' by Schmitz (2010) belonging to the *Drosophila bromeliae* group, from *Drosophila* genus) and *D. denieri* (belonging to the *Phloridosa* subgenus from *Drosophila* genus). Both species depend on flowers to develop their life cycle (Brncic, 1983; Schmitz, 2010). In our tests the lineages were maintained until F4 generation, in good performance.

The preparation of the medium uses common ingredients and the flowers can be kept in the freezer until their use, overcoming the natural periodicity of the blooms. Also, the equipment used is accessible making the routine preparation of this medium very cheap and easy.

Medium ingredients

0.22 g of agar
0.45 g of sugar
0.02 g of dry yeast biological
0.01 g of methylparaben
10ml of distilled water
1 macerated flower

Procedure

- Mix agar and 5 ml of distilled water and wait 10 min
- Add the remaining distilled water, the sugar, and the dry yeast
- Mix everything and boil on the microwave three times for 1 min

- Add the methylparaben and mix again
- Add the macerated flower and mix
- Transfer the medium to clean vials and let the medium cool protected from contaminants
- The use of a folded tissue in the medium is recommended

Comments

The flowers were collected and frozen at -10°C, or lower, in identified bags with flower species and collection date. The adults of P generation were collected with entomological aspirator in the mature flowers of *B. suaveolens* (used for *B. suaveolens* medium) and in *I. alba* (used for *I. alba* medium). The adults of P generation were placed in small vials containing the culture medium. These groups of flies are difficult to identify only using stereomicroscope. So, yellow adults (*Drosophila bromeliae* species group) were separated from black adults (*Phloridosa* subgenus species group). Adult females were placed separated in the medium for *B. suaveolens* and for *I. alba* and let oviposit for 5 days. After that, the P generation females were stored in 90% ethanol. Adult males, from all generations, were identified through the analysis of the terminalia using the technique from Wheeler and Kambyssellis (1996), modified by Bächli *et al.* (2004). After the first day of transference of the adults to a new medium, it was placed two drops of the flower extract prepared with 5 ml of distilled water, to feed the larvae and adults. All media were kept in a temperature and humidity controlled chamber ($\pm 25^{\circ}\text{C}$, 60% r.h.) reaching to the F4 generation in both kinds of media (*B. suaveolens* and *I. alba*) in good performance.

Acknowledgments: The authors thank to João Henrique Figueredo Oliveira for stimulus to develop this medium and to CNPq for fellowship and research grants.

References: Bächli, G., 2017, TaxoDros: The database on taxonomy of drosophilidae. Available at: <http://www.taxodros.uzh.ch/>. Accessed in: Dec 30th 2017; Bächli, G., C.R. Vilela, S.A. Escher, and A. Saura 2004, Fauna Entomologica Scandinavica; Brncic, D., 1983, In: *Genetics and Biology of Drosophila* vol. 3d. (Ashburner, M., H.L. Carson, and J.N. Thompson, jr., eds.). Academic Press; Markow, T.A., and P.M. O'Grady 2006, *Drosophila: A Guide to Species Identification and Use*. Elsevier; Marques, E.K., M. Napp, H. Winge, and A.B. Cordeiro 1966, Dros. Inf. Serv. 41: 187; Schmitz, H.J., 2010, Genética, Ecologia e Evolução de drosofilídeos (Insecta, Diptera) associados a flores. PhD thesis in Genetics and Molecular Biology, Universidade Federal do Rio Grande do Sul, Brazil.

Guide to Authors

Drosophila Information Service prints short research, technique, and teaching articles, descriptions of new mutations, and other material of general interest to *Drosophila* researchers. The current publication schedule for regular issues is annually, with the official publication date being 31 December of the year of the issue. The annual issue will, therefore, include material submitted during that calendar year. To help us meet this target date, we request that submissions be sent by 15 December if possible, but articles are accepted at any time. Receipt by 31 December is a firm deadline, due to printer submission schedules.

Manuscripts, orders, and inquiries concerning the regular annual DIS issue should be sent to James Thompson, Department of Biology, University of Oklahoma, Norman, OK 73019. Telephone (405)-325-2001; email jthompson@ou.edu; FAX (405)-325-7560.

Submission: Manuscripts should be submitted in Word, with pictures preferably in *.jpg. To help minimize editorial costs, proofs will not be sent to authors unless there is some question that needs to be clarified or they are specifically requested by the authors at the time of submission. The editor reserves the right to make minor grammatical, spelling, and stylistic changes if necessary to conform to DIS format and good English usage. Color illustrations will appear black and white in the printed version but will be in color in the electronically-accessible version on our web site (www.ou.edu/journals/dis).